HELIANGOLIDES FROM LEUCANTHEMOPSIS PULVERULENTA

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Abstract—The structure of hispanolide (4,5-cis-3-oxogermacranolide) and three other related germacranolides, isolated from *Leucanthemopsis pulverulenta*, have been established by a series of chemical transformations and spectral data.

INTRODUCTION

Leucanthemopsis (Anthemideae, Compositae) is a small genus, with only six species, most of them widespread in Spain and Portugal [1]. They have not previously been studied chemically.

In a previous paper [2], we reported the isolation of two new phloroglucinol derivatives from the roots of *L. pulverulenta*, previously known as *Pyrethrum hispanicum**, and now we report the isolation of six sesquiterpene lactones from the chloroform extract of the aerial parts. Four of these, 1-4, have not been reported previously; the other two lactones are the already known desacetyllaurenobiolide (5) and spiformin (6) [3].

RESULTS AND DISCUSSION

The new lactones are all structurally related heliangolides for which we propose the structure 4,5-cis-3-oxo-

*The plant was identified by Professor B. Casaseca Mena, from the Department of Botany, Salamanca University, where a specimen is held (Herbarium No. 8996).

germacranolide (1, hispanolide), 4.5-cis- 3β -acetoxygermacranolide (2), 4.5-cis-1.10-epoxy- 3β -acetoxygermacranolide (3) and 4.5-cis-1.10-epoxy- 3β -hydroxygermacranolide (4).

Hispanolide (1), mp 130–131°, showed a molecular ion at m/z 246, which agreed with the formula $C_{15}H_{18}O_3$. The UV spectrum, with λ_{\max}^{EIOH} at 213 ($\varepsilon=6.100$) and 254 ($\varepsilon=3.100$) nm, suggested the presence of an α -methylene- γ -lactone ring [4, 5] and a conjugated ketone with a near scis geometry [6]. This functionality was confirmed by the IR spectrum, which had bands at 1765, 1705 [7, 8], 1670, 1640 and 830 cm⁻¹. The ¹H NMR spectrum of 1 (Table 1) was very similar to that exhibited by 3-dehydronobilin (7) [9], the only significant difference being the additional ester signal at C-8 of the latter. These facts and the ¹H NMR coupling constants $J_{5,6}$, $J_{6,7}$ [10] and $J_{7,13}$ [11] (Table 1), allowed us to assign a heliangolide structure for 1.

The most probable conformation for hispanolide, deduced from these data and from analysis of the Dreiding models, is shown in A.

The ¹H NMR chemical shift of H-6 (δ 4.18) is due to the spatial proximity of the oxygen atom at C-3. That the

$$X$$
 $\frac{10^{9}}{14^{6}}$
 $\frac{10^{9}}{15^{13}}$
 $\frac{10^{9}}{15^{13}}$

1 X=0; R=H

1a X = OH, H; R = H

2 X = OAc, H; R = H

7 X = O; R = Ang

3 R = Ac

5

6

	1	2	3	4
H-1	5.32 t	5.20 m	2.78 dd	2.78 dd
	$J_{1,2} = 9 \text{ Hz}$		$J_{1,2} = 11, J_{1,2} = 4 \text{ Hz}$	$J_{1,2} = 11, J_{1,2} = 4 \text{ Hz}$
H-2	3.17 d			
	$J_{1.2} = 9 \text{ Hz}$			
H-3		5.15 t	5.35 t	4.45 t
		$J_{2.3} = 3 \text{ Hz}$	$J_{2.3} = 3 \text{ Hz}$	$J_{2,3} = 3 \text{ Hz}$
H-5	5.20 dd	5.12 dd	5.32 dd	5.25 dd
	$J_{5.6} = 11, J_{5.15} = 1.5 \text{ Hz}$	$J_{5.6} = 11, J_{5.15} = 1.5 \mathrm{Hz}$	$J_{5.6} = 11, J_{5.15} = 2.5 \text{ Hz}$	$J_{5.6} = 11, J_{5.15} = 1.5 \text{ Hz}$
H-6	4.18 dd	5.30 dd	5.66 dd	6.22 dd
	$J_{5.6} = 11, J_{6.7} = 2.5 \text{ Hz}$	$J_{5.6} = 11, J_{6.7} = 2.5 \text{ Hz}$	$J_{5.6} = 11, J_{6.7} = 2.5 \mathrm{Hz}$	$J_{5,6} = 11, J_{6,7} = 2.5 \text{ Hz}$
H-7	2.70 m	2.65 m	2.70 m	2.70 m
H-13	5.70 d	5.65 d	5.70 d	5.65 d
	$J_{7.13} = 2.5 \mathrm{Hz}$	$J_{7.13} = 2 \text{ Hz}$	$J_{7.13} = 2.5 \mathrm{Hz}$	$J_{7,13} = 2 \text{ Hz}$
H-13'	6.31 d	6.30 d	6.38 d	6.30 d
	$J_{7.13} = 2.5 \text{ Hz}$	$J_{7.13'} = 2 \text{ Hz}$	$J_{7.13'} = 2.5 \text{ Hz}$	$J_{7,13'} = 2 \text{ Hz}$
H-14	1.72 br s	1.71 br s	1.40 br s	1.41 s
H-15	1.88 d	1.75 d	1.82 d	1.75 d
	$J_{5,15} = 1.5 \text{ Hz}$	$J_{5.15} = 1.5 \mathrm{Hz}$	$J_{5.15} = 1.5 \text{ Hz}$	$J_{5.15} = 1.5 \text{ Hz}$

Table 1. ¹H NMR spectra of the lactones from Leucanthemopsis pulverulenta (60 MHz, δ, CDCl₃, TMS)

$$\Phi_{R} > 120^{\circ}$$

$$C_{8}$$

$$H$$

$$C_{5}$$

C=O group is not coplanar with the double bond accounts for the partial absence of conjugation (1705 cm⁻¹).

The structure and stereochemistry assigned to hispanolide was confirmed by reduction of 1 to 4.5-cis- 3β -hydroxygermacranolide (1a), of mp 137-138°, whose spectral data were identical with those exhibited by a germacranolide isolated by Bohlmann et al. [12] from Tanacetum tanacetoides. The ¹H NMR spectrum of 1a was very similar to that of hispanolide, but for the presence of a new allylic β -O-proton signal at δ 4.46 (t, J = 3 Hz) and the downfield shift of the H-6 signal (δ 5.80, dd, J = 11 and 2.5 Hz).

Acetylation of 1a, gave a monoacetate identical with the naturally occurring lactone 2. The ¹H NMR spectral data (Table 1), further support the proposed structure.

4,5-cis-1,10-Epoxy-3β-acetoxygermacranolide (3) was a gum showing a molecular ion at m/z 306, which agreed with the formula $C_{17}H_{22}O_5$. Its IR spectrum suggested the presence of an α-methylene-γ-lactone ring (1770 cm⁻¹), an acetoxyl group (1745 cm⁻¹) and an oxirane ring (1220 and 850 cm⁻¹). The structure was deduced from the characteristic ¹H NMR spectrum, which was very similar to that exhibited by 2, except for the upfield shift of the H-13 resonance from δ1.71 to 1.40 and that of the H-1 resonance from δ5.20 to 2.78. Hence, 3 is the epoxide of 2. This was confirmed by epoxidation of 2 with m-CPB which gave a substance identical in all respects with lactone 3. 4,5-cis-1,10-Epoxy-3β-hydroxy-

germacranolide (4), mp $168-169^{\circ}$, showed a molecular ion at m/z 264, which agreed with formula $C_{15}H_{22}O_4$.

The ¹H NMR spectrum of 4 was like that of 3, except for the absence of the upfield shift of the H-3 resonance, which indicated that the C-3 hydroxyl was free. In 4,5-cis-1,10-epoxy-3 β -hydroxygermacranolide (4) the H-6 signal (δ 6.22) was more deshielded than in any of the other hispanolide derivatives. This is due to the anisotropy of the three-membered oxirane ring, which has also been observed in other heliangolides [10]. Acetylation of 4, gave a monoacetate identical with 3.

The CD curves for hispanolide and its derivatives indicated that their absolute configurations are those depicted in structures 1–4 [9, 13–15].

EXPERIMENTAL

Mps are uncorr, and were determined on a Kofler hot stage apparatus.

Collection and isolation of plants. L. pulverulenta was collected at the end of May 1980, near Ledesma (Salamanca, Western Spain).

The aerial parts (2.3 kg) were air-dried, finely ground and extracted with hot CHCl₃. The extract was coned *in vacuo* and the residual syrup extracted with 50% aq. EtOH. The ethanolic soln was extracted with CHCl₃, and the CHCl₃ extract was then extracted with aq. 5% NaHCO₃, yielding a neutral fraction (6.2 g). The ethanolic soln after acidification with HCl, gave an acidic fraction (4.3 g).

The neutral fraction was dry chromatographed, using C_6H_6 –Et $_2O$ (7:3), to afford five fractions. Each of them was rechromatographed on Si gel or Si gel impregnated with 20% AgNO $_3$ and purified by crystallization or TLC. Fraction 1 (0.60 g) afforded 2 (200 mg). Fraction 2 (1.25 g) afforded 1 (82 mg) and 5 (340 mg). Fraction 3 (1.53 g) afforded 3 (240 mg) and 6 (50 mg). Fraction 4 (1.42 g) afforded 4 (535 mg). Fraction 5 (1.25 g) was a complex mixture.

Hispanolide (1). Compound 1 was eluted with CHCl₃–Et₂O (95:5); mp 130–131° (CHCl₃–Et₂O) and $[\alpha]_D = +148°$ (c 0.47, CHCl₃); UV λ_{\max}^{EIOH} nm: 213 (ε = 6.100) and 254 (ε = 6.100) nm; IR ν_{\max}^{KBr} cm⁻¹: 1765, 1705, 1670, 1640, 1340, 1280, 1150, 1030, 980, 940, 890, 860, 830; MS m/z (rel. int.): 246 [M]⁺ (5), 231 (7), 203 (16), 175 (28), 161 (100), 147 (72), 135 (33), 122 (31), 105 (94), 69 (43), 43 (58); CD: $\Delta \epsilon_{215} = -1.18$; $\Delta \epsilon_{305} = +1.12$.

Reduction of 1. To a soln of 1 (23 mg) in MeOH (3 ml), NaBH₄ (16 mg) in MeOH (2 ml) was added. The reaction mixture was kept at room temp. for 1 hr. The MeOH was evaporated, H₂O and two drops of HOAc were added and the organic phase was extracted with CHCl₃. Evaporation of the solvent gave 22 mg 1a, mp 137-138° (hexane-Et₂O), $[\alpha]_D = -18.1^\circ$ (c 0.43, CHCl₃).

Acetylation of 1a (14 mg) gave a monoacetate (13 mg) identical with 2.

4,5-cis-3 β -Acetoxygermacranolide (2). Compound 2 was eluted with C_6H_6 as a gum, $[\alpha]_D=-21^\circ$ (c 0.45, CHCl₃); IR ν_{max} cm $^{-1}$: 1770, 1740, 1670, 1460, 1380, 1250, 1190, 1140, 1050, 970, 890, 820; MS m/z (rel. int.): 290 $[M]^+$ (5), 248 $[M-42]^+$ (4), 230 $[M-60]^+$ (10), 149 (19), 121 (26), 107 (30), 93 (32), 67 (32), 43 (100); CD: $\Delta\epsilon_{212}=-3.48$; $\Delta\epsilon_{233}=+0.56$.

4,5-cis-1,10-Epoxy-3 β -acetoxygermacranolide (3). This was eluted with CHCl₃-Et₂O (9:1), $[\alpha]_D = -47^\circ$ (c 1.70, CHCl₃); IR ν_{max} cm⁻¹: 1770, 1750, 1660, 1450, 1380, 1250, 1140, 1090, 990, 890, 825, 805; MS m/z (rel. int.): 306 [M]⁺ (3), 264 [M - 42]⁺ (17), 246 [M - 60]⁺ (58), 218 (39), 203 (52), 175 (62), 159 (44), 147 (56), 135 (95), 123 (100), 122 (91), 95 (91), 43 (86).

Epoxidation of 2. To a stirred soln of 2 (17 mg) in CH_2Cl_2 (2 ml) at 0° , m-CPBA (8.6 mg) in CH_2Cl_2 (2 ml) was added and the mixture stirred for 30 min at 25°. A 10% aq. soln of Na_2SO_3 was added dropwise and the organic layer was washed \times 3 with a

5% aq. soln of Na₂CO₃, dried over Na₂SO₄ and evaporated to give 16 mg of 3.

4,5-cis-1,10-Epoxy-3 β -hydroxygermacranolide (4). Compound 4 was eluted with CHCl₃-Et₂O (8:2), mp 168–170° (Et₂O), [α]_D = -23.5° (c 1.8, CHCl₃); IR ν_{max} cm⁻¹: 3600–3100, 1765, 1670, 1450, 1380, 1280, 1140, 970, 880, 860, 820; MS m/z (rel. int.): 264 [M]⁺ (3), 246 [M - 18]⁺ (9), 231 (3), 203 (7), 175 (14), 161 (14), 147 (19), 135 (15), 123 (24), 95 (100), 91 (62), 69 (55), 55 (45); CD: $\Delta \varepsilon_{211} = -6.85$; $\Delta \varepsilon_{235} = +1.68$.

Acetylation of 4 (22 mg) gave a monoacetate (20 mg) identical with 3.

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